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Abstract

Background: Lujo virus (LUJV) is a highly fatal human pathogen belonging to the Arenaviridae family. Lujo virus causes viral hemorrhagic fever (VHF). An In silico molecular docking was performed on the GPC domain of Lujo virus in complex with the first CUB domain of neuropilin-2. The aim of this study is to predict an effective epitope-based vaccine against the glycoprotein GPC precursor of Lujo virus using immunoinformatics approaches. Methods and Materials: A glycoprotein GPC precursor of Lujo virus Sequence was retrieved from NCBI. Different prediction tools were then used to analyze the nominee's epitopes in BepiPred-2.0: Sequential B-Cell Epitope Predictor for B-cell, T-cell MHC class II I. Later the proposed peptides were docked using the Autodock 4.0 software program.

Index terms—

1 INTRODUCTION

Arenaviruses are rodent-borne viruses. Where a genetically unique arenavirus called Lujo virus, has been discovered as the causal agent of a nosocomial outbreak of acute febrile disease with hemorrhagic manifestations in Zambia and South Africa. The outbreak marked a high case fatality rate of almost 80% [1]. These viruses are genetically and geographically related to the Old World mammarena viruses, endemic to West Africa, and the New World mammarena viruses, endemic to South and North America [2]. Lujo virus causes viral hemorrhagic fever (VHF) which can be caused by five distinct families of viruses: the filo-, arena-, flavi-, rhabdo- and bunya virus family [3]. Viral hemorrhagic fever (VHF) is an acute systemic illness classically involving fever, a constellation of initially nonspecific signs and symptoms, and a propensity for bleeding and shock.

With Lujo virus hemorrhagic fever (LVHF) illness typically begins with the abrupt onset of fever, malaise, headache, and myalgias followed successively by a sore throat, chest pain, gastrointestinal symptoms, rash, minor hemorrhage, subconjunctival injection, and neck and facial swelling over the first week of sickness [4]. No major hemorrhage was noted. Whereas neurological signs were sometimes seen in the late stages, shock and multi-organ system failure, often with evidence of disseminated intravascular coagulopathy, ensued in the second week, with death in four of the five cases [4].

There are currently limited preventative and therapeutic options for patients infected with these highly pathogenic viruses [5].

Arenaviruses are enveloped negative-strand RNA viruses with a genome that is bi-segmented into S and L segments. The S segment encodes a nucleocapsid protein (NP) and an envelope glycoprotein precursor (GPC); the L segment encodes a matrix protein (Z) and an RNA-dependent RNA polymerase (L). The GPC is synthesized as a single polypeptide and undergoes processing by the host cell signal peptidase (SP ase) and subtilisin-like kexinisozyme-1/site-1 protease (SKI-1/S1P), yielding typical receptor binding (G1), transmembrane fusion (G2), and stable signal peptide (SSP) subunits, respectively [6-8]. Viral entry into target cells is initiated by the binding of G1 to appropriate cell surface receptors. The first cellular receptor for arenavirus to be identified was α -dystroglycan (α -DG), a ubiquitous receptor for extracellular matrix proteins [9]. The understanding of epitope/antibody interaction is the key to constructing potent vaccines and effective diagnostics. The host defense mechanisms against viruses generally vary from germline-encoded immunity, which present early in the evolution of microorganisms to activation and induction of specific adaptive immune responses by the production of Th-1 and Th-2 cytokines. B-cells recognize antigens via membrane bound antibodies using B-cell receptors (BCRs),

6 T CELL EPITOPE PREDICTION TOOLS 2.4.1 PEPTIDE BINDING TO MHC CLASS I MOLECULES

46 resulting in the secretion of antibodies that bind to the antigen and deactivate or remove it. Processing and
47 presentation of peptide epitopes are essential steps in cell-mediated immunity [10]. Lujo virus (LUJV) is a highly
48 fatal human pathogen belonging to the Arenaviridae family. This virus is unique; as it uses neuropilin-2 (NRP2)
49 as a cellular receptor.

50 Previous study revealed that the GP1 receptor-binding domain of LUJV (LUJVGPI) recognizes NRP2, where
51 its recognition is metal-ion dependent. The binding of a Ca^{2+} ion stabilizes the conformations of Asp127 and
52 Glu79 from NRP2 pre-organizing them for interaction with Lys110 of LUJVGPI. CUB domain of NRP2 is
53 almost completely conserved among humans, mice, rats and bats, and the only slight variations occur outside of
54 the binding site for LUJV. Hence all of these animal species have a potential to serve as reservoirs for LUJV,
55 considering only the compatibility to NRP2 [2]. In silico molecular docking was performed on the GP1 domain
56 of Lujo virus in complex with the first CUB domain of neuropilin-2 [2]. The aim of the study is to predict an
57 effective epitope-based vaccine against an envelope glycoprotein precursor (GPC) of Lujo virus. The development
58 of immunogenetics approaches will enhance the understanding of the genetic factors impact on the interindividual
59 and interpopulation variations in immune responses to vaccines that could be helpful to progress new vaccine
60 strategies [11]. In silico/reverse vaccinology had replaced conventional culture-based vaccine because it reduces
61 the cost required for laboratory investigation of pathogen, also speeding up the time needed to achieve the results
62 [12,13].

63 Therefore, using immunoinformatics approaches to predict this new kind of vaccines could be a London Journal
64 of Research in Computer Science and Technology magnificently additive in the way forward of preventing Lujo
65 virus. Normally, the investigation of the binding affinity of antigenic peptides to the MHC molecules is the
66 main goal when predicting epitopes. The usage of such tools and information leads to the development of new
67 vaccines. While these approaches permit the optimization of a London Journal of Research in Computer Science
68 and Technology vaccine for a specific population, it's probably can be reformulated to design a "universal vaccine"
69 a vaccine that provides maximum coverage for the whole world's population [14][15][16][17]. In this study, we
70 focused on both MHC class II and class I with performing of molecular docked in HLA-A0201.

71 2 II. MATERIALS AND METHODS

72 3 Sequences Retrieval

73 The amino acids sequences of Glycoprotein GPC (Glycoside hydrolase family) of Lujo virus were retrieved from
74 the NCBI database (<https://www.ncbi.nlm.nih.gov/protein>) [18] in FASTA format on July 2018. Different
75 prediction tools of Immune Epitope Database IEDB analysis resource (<http://www.iedb.org/>) [19] were then
76 used to analyze the candidate epitopes as shown on figure (1).

77 4 Conservation Region and Physicochemical Properties

78 Conservation regions were determined using multiple sequence alignment with the help of Clustal-W in the
79 BioEdit software version 7.2.5 [20]. Epitope conservancy prediction for individual epitopes was then calculated
80 using the IEDB analysis resource. Conservancy can be defined as the portion of protein sequences that restrain
81 the epitope measured at or exceeding a specific level of identity [21]. The physicochemical properties of the
82 retrieved sequence; molecular weight and amino acid composition; were also determined by BioEdit software
83 version 7.2.5 [20].

84 5 B Cell Epitope Prediction Tools

85 Candidate epitopes were analyzed using several B-cell prediction methods to determine their antigenicity,
86 flexibility, hydrophilicity, and surface accessibility. The linear prediction epitopes were obtained from the Immune
87 epitope database (<http://tools.iedb.org/bcell/result/>) [22] by using the BepiPred test with a threshold value of
88 0.149 and a window size of 6.0. Moreover, surface-accessible epitopes were predicated with a threshold value of
89 1.0 and window size of 6.0 using the Emini surface accessibility prediction tool [23].

90 Kolaskar and Tongaonker antigenicity methods (<http://tools.iedb.org/bcell/result/>) were proposed to deter-
91 mine the sites of antigenic epitopes with a default threshold value of 1.030 and a window size of 6.0 [24].

92 6 T cell epitope prediction tools 2.4.1 Peptide binding to MHC class I molecules

94 The peptide binding was assessed by IEDB MHC class I prediction tool at <http://tools.iedb.org/mhc1>. This
95 tool employs different methods to determine the ability of submitted sequence to bind to a specific MHC class
96 I molecule. The artificial neural network (ANN) method [25,26] was used to calculate IC₅₀ values of peptide
97 binding to MHC-class I molecules. For both frequent and non-frequent alleles, peptide length was set to 9
98 amino acids earlier to the prediction. The alleles having binding affinity IC₅₀ equal to or less than 500 nM were
99 considered for further analysis. The affinity of 500 nM is routinely used as a threshold for peptide selection and
100 it captures 92% of the epitopes [27][28][29].

101 7 Peptide Binding to MHC Class II Molecules

102 MHC class II prediction tool <http://tools.iedb.org/mhcII> provided by Immune Epitope Database (IEDB)
103 analysis resource and human allele references set was used to predict the peptide binding to MHC class II
104 molecules. Where the Artificial Neural Network prediction method was chosen to identify the binding affinity to
105 MHC class II grooves and MHC class II binding core epitopes. All epitopes that bind to as many alleles at score
106 equal to or less than 1000 half-maximal inhibitory concentration (IC₅₀) were selected for further analysis. [30] .

107 8 Population Coverage

108 Population coverage for each epitope was calculated by the IEDB population coverage tool at
109 http://tools.iedb.org/tools/population/iedb_input [31] . This tool was targeted in order to determine
110 the fraction of individual alleles predicted to respond to a given set of epitopes with known MHC restrictions.
111 For every single population coverage, the tool computed the following information: molecules were assessed
112 against a population coverage area selected prior to the submission.

113 9 Homology Modeling

114 The 3D structure of glycoprotein GPC of Lujo virus was predicted using Raptor X web portal
115 (<http://raptorx.uchicago.edu/>) [32] . The reference sequence was submitted in FASTA format on 14/9/2018 and
116 the structure was received on 15/9/2018. Subsequently the structure was treated with UCSF Chimera 1.10.2
117 [33] to visualize the position of the proposed peptides.

118 10 In Silico Molecular Docking 2.7.1 Ligand Preparation

119 In order to estimate the binding affinities between the epitopes and the molecular structure of MHC class I &
120 MHC class II, in silico molecular docking was utilized. The proposed epitopes sequences were then selected from
121 the Lujo virus reference sequence using Chimera 1.10 and saved as a "pdb" file. The obtained files were later
122 optimized and energy minimized. The HLA-A0201 was selected as the macromolecule for docking; as HLA-A0201
123 is considered as the most popular MHC allele and most MHC-I epitopes were nonapeptides [34] .

124 Its crystal structure (4UQ3) was downloaded from the RCSB Protein Data Bank (<http://www.rcsb.org/pdb/home/home.do>), which was in complex with an azobenzene-containing peptide [35] .

125 The crystal structure of LUJVGP1/NRP2 was retrieved from protein databank (PDB ID: 6GH8) [2] .

127 11 Molecular Docking

128 Molecular docking was performed using Autodock 4.0 software, based on Lamarckian Genetic Algorithm; which
129 combines energy evaluation through grids of affinity potential to find the suitable binding position for a ligand
130 on a given protein [36] . Polar hydrogen atoms were added to the protein targets and Kollman united atomic
131 charges were computed. All hydrogen atoms were added to the ligands before the Gastiger partial charges were
132 assigned. The co-crystal ligand was removed and the bond orders were checked. The target's grid map was
133 calculated and set to 60×60×60 points with a grid spacing of 0.375 Å. The grid box was then allocated properly
134 in the target to include the active residue in the center. The default docking algorithms were set in accordance
135 with standard docking protocol [37] . Finally, ten independent docking runs were carried out for each ligand
136 and results were retrieved as binding energies. Poses that showed the lowest binding energies were visualized
137 using the UCSF chimera [38] .

138 III. RESULTS

139 12 Lujo Virus Glycoprotein GPC Physical and Chemical Parameters

140
141 The physicochemical properties of the Lujo virus glycoprotein GPC protein was assessed using BioEdit software
142 version 7.0.9.0. The protein length was found to be 454 amino acids. The amino acids that form Lujo virus
143 glycoprotein GPC protein is shown in Figure (2) along with their numbers and molar percentages in (Mol%).

144 13 Prediction of T Helper Cell Epitopes and Interaction With MHC Class I Alleles

145
146 Lujo virus glycoprotein GPC sequence was analyzed using IEDB MHC class I binding prediction tool based on
147 ANN-align with half-maximal inhibitory concentration (IC₅₀) >500; the least most promising epitopes that had
148 a binding affinity with the Class I alleles along with their positions in the Lujo virus glycoprotein GPC are shown
149 in Table ??.

14 Prediction of T Helper Cell Epitopes and Interaction With MHC Class II Alleles

Lujo virus glycoprotein GPC sequence was analyzed using IEDB MHC class II binding prediction tool based on NN-align with half-maximal inhibitory concentration (IC₅₀) >1000. The list of the most promising epitopes, that had a strong binding affinity to MHC class II alleles and depending on the number of their binding alleles are shown in Table ??.

15 Table 3: Most potential T-cell Epitopes with Interacting MHC-Class II Alleles

16 IV. DISCUSSION

In this computational immunoinformatic study we suggest a new promising highly selective peptides vaccine against Lujo virus for the first time according to our findings. We expect to obtain a peptide-based vaccine which implies a high antigenicity and a minimum allergic outcome that is more accurate than the currently used vaccines. The analytical process started after having adequate information on the protein structure of Lujo virus according to the literature review. Simultaneously, though the 3D structure was previously available on the database with all its prospects, we produced our own structure using Raptor X web portal to utilize its complete physicochemical properties information file to confirm our results, and it's a technique we have pursued. The reference sequence of Lujo virus glycoprotein GPC was obtained from the NCBI database. To determine the binding affinity of the conserved epitopes to B-cell and to examine the immunogenicity several tests on the IEDB database were used; the Bepipred linear epitope prediction test, Emini surface accessibility test, and Kolaskar and Tongaonkar antigenicity test were examined. For the Bepipred test of B-cell, the total number of epitopes was 39. For Emini surface accessibility prediction, 29 conserved epitopes were passing the default threshold of 1.0.

In Kolaskar and Tongaonkar antigenicity, 7 epitopes provided a score above the default threshold 1.045. However, there are only two epitopes that passed our three tests, which were (LPCPKPHRLR, LPCPKPHR). The reference glycoprotein GPC strain was analyzed using IEDB class MHC-class I binding prediction tool to predict T cell epitope. According to these interesting findings, a very promising vaccine against Lujo virus can potentially be formulated. The most promising three peptides; FWYLNHTKL, LPCPKPHRLR and YMFSVTLCI were docked on to protein target of GPC domain of Lujo virus in complex with the first CUB domain of NRP2 as shown on figure (4 -6). All peptides were docked on the interface of Lujo virus GPC/NRP2 and scored binding energies of -5.84, -3.88 and -8.20 Kcal/mol for peptides 1, 2 and 3, respectively. As the docking results from the analysis of peptide-1 showed binding hydrogen bonding with two amino acid residues: SER-51 and GLN-131 of NRP2. While, Peptide-2 showed a bonding affinity to hydrogen with residues HIS-131 and PHE-137 of Lujo virus GPC and ARG-432 of NRP2, whereas peptide-3 formed hydrogen bonds with PHE-137 of Lujo virus GPC.

Only Peptide-2 and 3 interact by forming hydrogen bonding with residues on Lujo virus GPC (HIS-131 and PHE-137). These residues are located at a hydrophobic pocket and adjacent to both residues Val139 and Thr140 of the ?2?4 loop which participates in Van der Waals interactions with NRP2 residues. In addition, histidine residues in the Lujo virus GPC/NRP2 complex are obvious candidates for controlling pH-dependent protein-protein interactions.

As for peptide-1, it has formed hydrogen bonds with GLN-131 of NRP2, which is adjacent to the key residue Arg130 that is important for NRP2-fc to recognize Lujo virus GPC-bearing cells and cell entry of Lujo virus [2]. The overall docking results analysis has revealed that the peptides are docked at Lujo virus GPC/NRP2 binding surfaces, in which these peptides would serve as potential inhibitors for blocking binding to NRP2 and thus may neutralize the virus. To the best of our knowledge, this study is considered to be the first to propose an epitope-based peptide vaccine against glycoprotein GPC of Lujo virus, which is expected to be highly antigenic with a minimum allergic impact. Furthermore, this study proposes a promising peptide FWYLNHTKL with a very strong binding affinity to MHC1 and MHC11 alleles. This peptide shows exceptional population coverage results for both MHC1 and MHC11 alleles.

In-vivo and in-vitro assessments for the most promising peptides namely, FWYLNHTKL, LPCPKPHRLR and YMFSVTLCI are recommended to be explored and studied on their ability to be developed into vaccines against Lujo virus glycoprotein GPC.

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² Epitope -based Peptide Vaccine Against Glycoprotein GPC Precursor of Lujo Virus using Immunoinformatics Approaches

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1

Figure 1: Figure 1 :



Figure 2:

2

Figure 3: Figure 2 :



2

Figure 4: Table 2 :



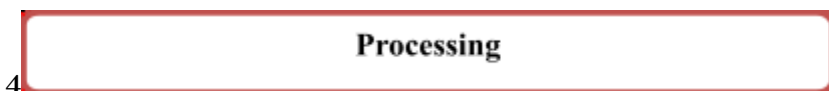
135

Figure 5: 13 3. 5



327

Figure 6: Figure 3 : 2 3. 7



4

Figure 7: Figure 4 :



Figure 8: Figure 5 :



Figure 9: Figure 6 :

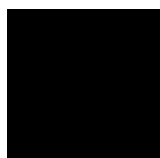


Figure 10:

1

No.	Start	End	Peptide	Length	Emini Surfce	scor	Kolasker & Tongankar	scor
33	423	432	LPCPKP	10	pass	1.371	pass	1.088
35	423	430	HRLR LPCPKP	8	pass	1.378	pass	1.095
			HR					

Figure 11: Table 1 :

4

MHC classes	Population	Parts of the World		
		World	South Africa	Sudan
Class I	Coverage a	99.83%	99.4%	99.41%
	Average_hit b	32.5	25.45	28.25
	PC90 c	13.28	8.99	8.76
Class II	Coverage a	68.23%	32.1%	56.38%
	Average_hit b	55.22	10.89	34.14
	PC90 c	-6.57	4.71	5.57

a projected population coverage

b average number of epitope hits / HLA combinations recognized by the population

c minimum number of epitope hits / HLA combinations recognized by 90% of the population.

Figure 12: Table 4 :

5

Peptide	Population coverage %/ Area						
	World		South Africa		Sudan		
	MHC I	MHC II	MHC I	MHC II	MHC I & II	MHC I	MHC II
	56.92%	88.77%			5.91%	65.72%	21.85%
Ymfsvtlci	73.92%		63.56%				75.17%
	42.99%	74.82%	41.97%		1.79%	43.01%	35.12%
Fwylnhtkl55	84%						25.35%

Epitope -based Peptide Vaccine Against Glycoprotein GPC Precursor of Lujo Virus using Immunoinformatics Approaches

Figure 13: Table 5 :

5

to interact with different class MHC-class I alleles. For class MHC-class II binding prediction, there were 315 epitopes found to interact with class MHC-class II alleles. The peptides
 RLQEAVSTL,
 VIFDLFREF, ITFSLLTNK, ILMFSVSFY and FWYLNHTKL had the affinity to bind with the highest number of class MHC-class I alleles. The peptides
 FNMSLLSSI, INAIISDTL, LMKLFQWSL and VFQAIPEIL had the affinity to bind with the highest number of class MHC-class II alleles. The most promising three peptides for both class MHC-class I and MHC-class II were FWYLNHTKL, LPCPKPHRLR and YMFSVTLCI as shown on figure (3). On the other hand, the world Population coverage of all epitopes that bind to MHC-class I were found to be 99.83%, while the world population coverage of all epitopes that bind to MHC-class II were 68.23% as presented in table 4. For the binding affinity to MHC-class I and MHC-class II the peptide FWYLNHTKL was found to bind 14 different alleles of MHC-class II & five alleles of MHC-class I, that gave a world population coverage of 74.82% , 43.01% for South Africa and 51.56% for Sudan of both MHC class I and II as shown on

peptides were predicted

YMFSVTLCI, SFYFYM,
 FQLVNSLLSSIPM,

FWYLNHTKL, LCI,

Figure 14: table 5 .

1 Competing Interests

The authors declare that they have no competing interests.

[London Journal of Research in Computer Science and Technology] , *London Journal of Research in Computer Science and Technology*

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