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Margarita A. Sazonova, Natalya A. Doroschuk, Marina D. Sazonova, Anastasia I. Ryzhkova, Mikhail A. Popov, Vasily N. Sukhorukov, Tatiana I. Kovyanova, Irina A. Starodubtseva, Dmitry F. Belyoyartsev, Alexey V. Churov, Paolo Poggio, Donato Moschetta, Vasily P. Karagodin, Anton Yu. Postnov & Alexander N. Orekhov

NN Burdenko Voronezh State Medical University

ABSTRACT

One of the possible factors in the occurrence of coronary heart disease with myocardial infarction is oxidative stress. Oxidative stress intensifies the formation of free radicals, causing defects in proteins and nucleic acids. This can lead to their partial or complete destruction. For example, the result of such destruction of DNA molecules may be a decrease in the length of telomeric repeats in the chromosomes of cells in coronary heart disease.

Therefore, the aim of this work was to analyze the association of certain variants of the relative length of telomeric repeats (VRLTR) with coronary heart disease with old myocardial infarction (CHD with MI).

Keywords: relative length of telomeric repeats; coronary heart disease with myocardial infarction; percentage of the calibrator; oxidative stress; protective effect; chromosomes.

Classification: NLM Code: QU 475

Language: English



Great Britain
Journals Press

LJP Copyright ID: 392881

London Journal of Medical & Health Research

Volume 24 | Issue 7 | Compilation 1.0



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New Markers of Coronary Heart Disease with Old Myocardial Infarction: Certain Variants of Relative Length of Telomeric Repeats

Margarita A. Sazonova^a, Natalya A. Doroschuk^a, Marina D. Sazonova^b,
Anastasia I. Ryzhkova^c, Mikhail A. Popov^y, Vasily N. Sukhorukov^d, Tatiana I. Kovyanova^x,
Irina A. Starodubtseva^v, Dmitry F. Beloyartsev^e, Alexey V. Churov^c, Paolo Poggio^f,
Donato Moschetta^e, Vasily P. Karagodin^f, Anton Yu. Postnov^e & Alexander N. Orekhov^g

ABSTRACT

One of the possible factors in the occurrence of coronary heart disease with myocardial infarction is oxidative stress. Oxidative stress intensifies the formation of free radicals, causing defects in proteins and nucleic acids. This can lead to their partial or complete destruction. For example, the result of such destruction of DNA molecules may be a decrease in the length of telomeric repeats in the chromosomes of cells in coronary heart disease.

Therefore, the aim of this work was to analyze the association of certain variants of the relative length of telomeric repeats (VRLTR) with coronary heart disease with old myocardial infarction (CHD with MI).

To measure the length of telomeric repeats, whole blood was collected, with the following DNA isolation from nuclear cells. The relative length of telomeric repeats was calculated based on the formula “2 to the power (-ΔCt)”, where $\Delta Ct = Ct$ of telomeres - Ct of albumin. In this case, Ct of telomeres is the threshold cycle of the telomeric repeat, and Ct of albumin is the threshold cycle of the albumin gene. The results of the relative length of telomeric repeats are presented as a percentage of the calibrator. DNA isolated from HeLa cell line was used as a calibrator.

By measuring variants of the relative length of telomeric repeats in the studied samples, an association of coronary heart disease with myocardial infarction with VRLTR-46,

VRLTR-49, VRLTR-51, VRLTR-53 and VRLTR-56 was detected. At the same time, 6 of 21 variants of the relative length of telomeric repeats (VRLTR-63, VRLTR-65, VRLTR-68, VRLTR-70, VRLTR-71 and VRLTR-73) had a protective effect in coronary heart disease with myocardial infarction.

Therefore, in the present research work, we propose new markers of coronary heart disease with old myocardial infarction: certain variants of relative length of telomeric repeats.

Keywords: relative length of telomeric repeats; coronary heart disease with myocardial infarction; percentage of the calibrator; oxidative stress; protective effect; chromosomes.

Author a σ p C O § X ζ F G: Laboratory of Angiopathology, Institute of General Pathology and Pathophysiology, 8 Baltiiskaya Street, Moscow, 125315, Russia.

a ε: Laboratory of Medical Genetics, Institute of Experimental Cardiology, National Medical Research Center of Cardiology, 15a, 3rd Cherepkovskaya Str., Moscow, 121552, Russia.

§ ε G: Laboratory of Cellular and Molecular Pathology of Cardiovascular System, Federal State Budgetary Scientific Institution, Petrovsky National Research Centre of Surgery (FSBSI "Petrovsky NRCS"), Moscow, Russia.

X: Institute for Atherosclerosis Research, Osennyyaya Street 4-1-207, 121609, Moscow, Russia.

y: Department of Cardiac Surgery, Moscow Regional Research and Clinical Institute ("MONIKI"), 61/2, Shchepkin St., Moscow, 129110, Russia.

v: Department of Polyclinic Therapy, NN Burdenko Voronezh State Medical University, 10 Studencheskaya Street, 394036 Voronezh, Russia.

E: Vascular Surgery Department, A. V. Vishnevsky National Medical Research Center of Surgery, 27 Bolshaya Serpukhovskaya Street, 117997 Moscow, Russia.

ζ: Pirogov Russian National Research Medical University, Russian Gerontology Clinical Research Centre, Moscow, Institute on Aging Research, Russian Federation, 16 1st Leonova Street, 129226 Moscow, Russia.

F: Plekhanov Russian University of Economics, Stremyanny lane, 36, Moscow, 117997, Russia.

Ε, Ε: Unit for the Study of Aortic, Valvular and Coronary Pathologies Centro Cardiologico Monzino, IRCCS.Via C. Parea, 4, Milano, 20138, Italy.

I. INTRODUCTION

One of the possible factors in the occurrence of coronary heart disease with myocardial infarction is oxidative stress [1-5]. Oxidative stress intensifies the formation of free radicals, causing defects in proteins and nucleic acids [6-10]. This can lead to their partial or complete destruction [11-15]. For example, the result of such destruction of DNA molecules may be a decrease in the length of telomeric repeats in the chromosomes of cells in coronary heart disease [16-20].

However, to date, there is no reliable data confirming the association of certain variants of telomeric repeat lengths with coronary heart disease with old myocardial infarction.

Telomeres are regions of chromosomal ends which contain tandem six-nucleotide repeats (TTAGGG) [21-25]. It is believed that they are intended for stable cell replication [26-30]. With each subsequent cell mitosis, the size of telomeric repeats decreases. Evidence has been obtained that a critical decrease in telomere size can cause degenerative changes in the cell followed by apoptosis [31-35].

It has been noted that individuals who have very short telomeres have a higher risk of developing coronary heart disease [35-40]. Scientists from Scotland have shown that individuals with shortened telomeres have a threefold increased risk of death over the next decade [41]. Short telomeric repeat lengths have been found in

elderly patients with cardiomyopathy [42]. Type 2 diabetes mellitus has been found to be significantly increased in individuals with short telomeres compared to individuals with long telomeres [43].

Meanwhile, it was found that the speed of decrease in the length of telomeric repeats is almost the same for different types of cells and tissues. Therefore, the use of human blood leukocytes, as the most accessible biological material for research, seems to be optimal [42, 43].

So, the main direction in the study of telomeres turned out to be the detection of the association between certain variants of shortened telomeres and certain human diseases. For this reason, it seems to be appropriate to include the analysis of telomeric repeat length variants into the biomarkers studied in coronary heart disease with myocardial infarction.

II. MATERIALS AND METHODS

To measure the length of telomeric repeats, whole blood was collected, with the following DNA isolation from nuclear cells. The DNA extraction kit "DNA-Extran1" (CJSC Synthol, Russia) was used. The length of telomeric repeats was determined by quantitative real-time polymerase chain reaction (RT-PCR) using a BIO-RADCFX 96 Real-Time System amplifier (Singapore). The study of each sample was repeated three times.

The relative length of telomeric repeats was calculated based on the formula " $2^{-\Delta Ct}$ ", where $\Delta Ct = Ct_{\text{telomeres}} - Ct_{\text{albumin}}$. In this case, $Ct_{\text{telomeres}}$ is the threshold cycle of the telomeric repeat, and Ct_{albumin} is the threshold cycle of the albumin gene. It should be noted that the albumin gene acted as an internal control against which the length of telomeric repeats was determined. The results of the relative length of telomeric repeats are presented as a percentage of the calibrator. DNA isolated from HeLa cell line was used as a calibrator.

Since there were no significant age differences between the samples of patients with coronary

heart disease who had suffered a myocardial infarction and conditionally healthy study participants, normalization of the length of telomeric repeats by age was not performed.

Statistical analysis of the relative length of telomeric repeats was carried out using the IBM SPSS Statistics 27.0 software package.

III. RESULTS AND DISCUSSION

In 300 DNA samples from samples of patients with coronary heart disease with old myocardial infarction (CHD with MI) and conditionally healthy study participants, variants of the relative length of telomeric repeats (RTL) were analyzed using real-time PCR.

Methods of clinical examination, high-resolution B-mode ultrasonography and DNA extraction were used. The criteria for inclusion in the sample of patients CHD with MI were-

1. Men and women aged 45 to 79 years.
2. Old myocardial infarction in the anamnesis.
3. Pathological Q waves with or without symptoms in the absence of non-ischemic causes.
4. Lack of exclusion criteria.
5. Voluntary consent to participate in the study.

The criteria for excluding patients CHD with MI were:

1. Age younger than 45 or older than 79 years.
2. Abnormal anatomical configuration of the neck and muscles; pronounced tortuosity and/or depth of the carotid arteries, and/or unusual locations of arterial branches.
3. Presence of oncological and other chronic diseases requiring regular drug therapy, except arterial hypertension.

Table 1: Analysis of the Average Value of Variants of the Relative Length of Telomeric Repeats in the Blood Leukocytes of Study Participants

Samples of Study Participants	Mean Relative Length of Telomeric Repeats	Standard Error to the mean	Reliability of Results
Patients with coronary heart disease with old myocardial infarction	51	1.1	$p \leq 0.001$
Conditionally healthy study participants	68	1.4	$p \leq 0.05$

Table 2: Analysis of Variants of the Relative Length of Telomeric Repeats in Samples of Patients with Coronary Heart Disease with Old Myocardial Infarction (CHD With MI) and Conditionally Healthy Study Participants using the Wilcoxon Rank Test

Variants of the Relative Length of Telomeric Repeats	Type of Rank	Number of Ranks	Mean Rank	Sum rank
VRLTR -31	Negative	2	3.11	3.18
	Positive	2	1.27	1.65
	Neutral	3		
VRLTR -32	Negative	1	3.15	4.17
	Positive	2	2.56	4.58
	Neutral	4		
VRLTR -33	Negative	2	2.16	3.18
	Positive	2	2.05	3.16
	Neutral	3		
VRLTR -34	Negative	2	1.43	1.72
	Positive	1	2.26	2.26
	Neutral	4		
VRLTR -35	Negative	2	2.04	2.17
	Positive	2	1.12	1.21
	Neutral	3		
VRLTR-36	Negative	2	2.08	3.06
	Positive	3	3.14	4.08
	Neutral	2		
VRLTR-37	Negative	2	2.02	3.27
	Positive	2	3.02	4.09
	Neutral	3		
VRLTR-38	Negative	2	2.16	2.48
	Positive	3	2.29	2.87
	Neutral	2		
VRLTR-39	Negative	2	3.05	4.21
	Positive	1	4.83	4.83

	Neutral	4		
VRLTR-40	Negative	2	2.30	3.11*
	Positive	4	1.51	6.41*
VRLTR-41	Neutral	1		
	Negative	1	4.22	4.22*
	Positive	2	3.78	8.63*
	Neutral	3		
VRLTR-42	Negative	2	4.18	6.03
	Positive	2	3.05	4.97
	Neutral	3		
	Negative	2	2.75	3.18*
	Positive	4	3.21	6.69*
	Neutral	1		
VRLTR-44	Negative	1	2.19	2.19*
	Positive	2	2.87	4.81*
	Neutral	4		
	Negative	3	2.11	3.94
	Positive	1	3.16	3.16
	Neutral	3		
VRLTR-46	Negative	2	3.17*	7.30*
	Positive	4	5.32*	21.33*
	Neutral	1		
	Negative	1	3.98	3.98*
VRLTR-47	Positive	2	4.01	8.04*
	Neutral	3		
VRLTR-48	Negative	3	5.35	6.24
	Positive	2	4.14	6.45
	Neutral	2		
	Negative	2	1.10*	2.24*
	Positive	4	2.21*	8.96*

	Neutral	1		
VRLTR-50	Negative	4	3.11	5.03
	Positive	2	2.15	3.15
VRLTR-51	Neutral	1		
	Negative	2	2.08*	6.18*
VRLTR-52	Positive	4	6.16*	12.36*
	Neutral	1		
VRLTR-53	Negative	2	3.25	3.98
	Positive	1	4.08	4.08
VRLTR-54	Neutral	4		
	Negative	2	2.60*	7.84*
VRLTR-55	Positive	3	5.45*	16.75*
	Neutral	2		
VRLTR-56	Negative	4	6.71	7.56
	Positive	1	4.12	4.12
VRLTR-57	Neutral	2		
	Negative	2	1.50	2.11
VRLTR-58	Positive	2	2.15	2.94
	Neutral	3		
VRLTR-59	Negative	2	2.50*	5.09*
	Positive	4	5.04*	20.24*
VRLTR-57	Neutral	1		
	Negative	3	3.64	4.14
VRLTR-58	Positive	3	5.16	6.12
	Neutral	1		
VRLTR-59	Negative	3	5.16	6.28
	Positive	2	3.11	3.54
VRLTR-59	Neutral	3		
	Negative	4	2.28*	5.56
VRLTR-59	Positive	2	1.14*	4.27

	Neutral	1		
VRLTR-60	Negative	3	3.16	4.12
	Positive	2	5.41	5.98
	Neutral	1		
VRLTR-61	Negative	2	1.14	1.50*
	Positive	3	2.01	3.00*
	Neutral	2		
VRLTR-62	Negative	2	3.33	4.51
	Positive	2	2.50	3.57
	Neutral	3		
VRLTR-63	Negative	4	5.08*	20.46*
	Positive	2	2.24*	5.11*
	Neutral	1		
VRLTR-64	Negative	2	2.19	3.74
	Positive	2	3.11	4.26
	Neutral	3		
VRLTR-65	Negative	4	4.06*	16.28*
	Positive	2	2.02*	4.11*
	Neutral	1		
VRLTR-66	Negative	1	1.32	1.32
	Positive	2	1.54	2.18
	Neutral	4		
VRLTR-67	Negative	2	2.37	2.94
	Positive	2	3.68	4.01
	Neutral	3		
VRLTR-68	Negative	4	4.25*	17.48*
	Positive	2	2.16*	4.34*
	Neutral	1		
VRLTR-69	Negative	1	1.15	1.15
	Positive	2	1.34	1.68

	Neutral	3		
VRLTR-70	Negative	4	6.12*	25.56*
	Positive	2	2.11*	4.26*
	Neutral	2		
VRLTR-71	Negative	4	6.18*	26.83*
	Positive	2	2.01*	4.04*
	Neutral	1		
VRLTR-72	Negative	3	3.06	4.10
	Positive	2	2.34	3.72
	Neutral	2		
VRLTR-73	Negative	4	4.17*	16.71*
	Positive	2	2.08*	4.18*
	Neutral	1		
VRLTR-74	Negative	1	1.16	1.16
	Positive	2	1.19	1.82
	Neutral	3		
VRLTR-75	Negative	1	2.96	2.96
	Positive	1	3.76	3.76
	Neutral	3		
VRLTR-76	Negative	4	2.38	9.56*
	Positive	2	2.04	4.71*
	Neutral	1		
VRLTR-77	Negative	2	3.24	4.59
	Positive	1	3.22	3.22
	Neutral	3		
VRLTR-78	Negative	3	5.33	8.35
	Positive	3	5.62	7.68
	Neutral	1		
VRLTR-79	Negative	3	7.23	21.80*
	Positive	1	3.15	3.15*

	Neutral	3		
VRLTR-80	Negative	4	6.18*	8.81
	Positive	2	3.07*	5.94
	Neutral	1		

*Note: * is more than two-fold difference between positive and negative rank value.*

According to the Wilcoxon rank test, 21 variants of the relative length of telomeric repeats were detected to be associated with coronary heart disease with myocardial infarction: VRLTR-40, VRLTR-41, VRLTR-43, VRLTR-44, VRLTR-46, VRLTR-47, VRLTR-49, VRLTR-51, VRLTR-53, VRLTR-56, VRLTR-59, VRLTR-61, VRLTR-63, VRLTR-65, VRLTR-68, VRLTR-70, VRLTR-71, VRLTR-73, VRLTR-76, VRLTR-79 and VRLTR-80.

During a correlation analysis, for 5 of the 21 detected VRLTR variants, highly significant differences were found between samples of patients with CHD with MI and conditionally healthy study participants (Table 3). In particular, an association of coronary heart disease with old myocardial infarction with variants of the relative length of telomeric repeats VRLTR-46, VRLTR-49, VRLTR-51, VRLTR-53 and VRLTR-56 was identified. At the same time, 6 of 21 variants of the relative length of telomeric repeats (VRLTR-63, VRLTR-65, VRLTR-68, VRLTR-70, VRLTR-71 and VRLTR-73) had a protective effect in coronary heart disease with old myocardial infarction.

IV. CONCLUSION

In the present research work, we propose new markers of coronary heart disease with old myocardial infarction: certain variants of relative length of telomeric repeats (VRLTR). These markers in the studied samples were presented as percentages of the calibrator. DNA isolated from HeLa cells was used as a calibrator.

By measuring variants of the relative length of telomeric repeats in the studied samples, an association of coronary heart disease with myocardial infarction with VRLTR-46, VRLTR-49, VRLTR-51, VRLTR-53 and VRLTR-56

was detected. At the same time, 6 of 21 variants of the relative length of telomeric repeats (VRLTR-63, VRLTR-65, VRLTR-68, VRLTR-70, VRLTR-71 and VRLTR-73) had a protective effect in coronary heart disease with myocardial infarction.

Table 3: Correlation of Variants of the Relative Length of Telomeric Repeats with Coronary Heart Disease with Old Myocardial Infarction

Number	Variants of the relative length of telomeric repeats	Correlation coefficient	Asymptomatic significance (two-sided)
1	VRLTR-40	0.211	0.101*
2	VRLTR-41	0.302	0.064*
3	VRLTR-43	0.287	0.071*
4	VRLTR-44	0.218	0.096*
5	VRLTR-46	0.609	0.001**
6	VRLTR-47	0.305	0.068*
7	VRLTR-49	0.591	0.002**
8	VRLTR-51	0.610	0.001**
9	VRLTR-53	0.554	0.003**
10	VRLTR-56	0.621	0.001**
11	VRLTR-59	-0.312	0.056*
12	VRLTR-61	-0.306	0.068*
13	VRLTR-63	-0.608	0.001**
14	VRLTR-65	-0.615	0.001**
15	VRLTR-68	-0.425	0.043**
16	VRLTR-70	-0.554	0.036**
17	VRLTR-71	-0.482	0.031**
18	VRLTR-73	-0.542	0.042**
19	VRLTR-76	-0.244	0.076*
20	VRLTR-79	-0.267	0.071*
21	VRLTR-80	-0.326	0.054*

Note: * is highly significant correlation of mutations with CHD with MI ($p \leq 0.05$);

** is correlation of mutations with coronary heart disease with myocardial infarction at the level of significance $p \leq 0.1$.

Author Contributions

Conceptualization, M.A.S.; methodology, M.A.S.; validation, A.I.R. and M.D.S.; formal analysis, M.A.S. and M.D.S.; investigation, M.A.S., N.A.D., M.D.S. and A.I.R.; resources, A.Yu.P., T.I.K., I.A.S., D.F.B. and A.V.Ch.; data curation, M.A.S., N.A.D. and M.D.S.; Writing – Original Draft

Preparation, M.A.S., M.D.S., N.A.D., A.I.R., P.P. and D.M.; Writing – Review & Editing, V.P.K., M.A.P. and A.Yu.P.; Project Administration, A.N.O. and V.N.S; Funding Acquisition, A.N.O.

All authors have read and agreed to the published version of the manuscript.

Funding

This work was financially supported by the Russian Science Foundation, grant #24-15-00123 (creating samples, analyzing results) and the Ministry of Science and Higher Education of the Russian Federation, Project # FGFU-2022-00008 (writing and designing the article).

Institutional Review Board Statement

The study was performed in accordance with the principles outlined in the Declaration of Helsinki of 1975 and its revised version of 2013. The study protocol was approved by the Institute for Atherosclerosis Research Committee on Human Research, Moscow, Russia, protocol No. 078-15 of September, 08, 2015.

Informed Consent Statement

Informed consent was obtained from all subjects involved in the study.

Data availability statements

The datasets presented in this article are not readily available because the data are part of an ongoing study of patients with cardiovascular diseases from Russia.

ACKNOWLEDGEMENT

We thank the Laboratory of Medical Genetics of Chazov National Medical Research Center of Cardiology for the facilities and instruments provided.

Conflict of interests

The authors declare no conflict of interests. The material for the investigation has never been published before.

REFERENCES

- Cheng, S., Han, Y., Jiang, L., Lan, Z., Liao, H., & Guo, J. (2023). Associations of oxidative balance score and visceral adiposity index with risk of ischaemic heart disease: a cross-sectional study of NHANES, 2005–2018. *BMJ open*, 13(7), e072334. <https://doi.org/10.1136/bmjopen-2023-072334>.
- Bai, B., Ji, Z., Wang, F., Qin, C., Zhou, H., Li, D., & Wu, Y. (2023). CTRP12 ameliorates post-myocardial infarction heart failure through down-regulation of cardiac apoptosis, oxidative stress and inflammation by influencing the TAK1-p38 MAPK/JNK pathway. *Inflammation research: official journal of the European Histamine Research Society ... [et al.]*, 72(7), 1375–1390. <https://doi.org/10.1007/s00011-023-01758-4>.
- Fuentes, E., Moore-Carrasco, R., de Andrade Paes, A. M., & Trostchansky, A. (2019). Role of Platelet Activation and Oxidative Stress in the Evolution of Myocardial Infarction. *Journal of cardiovascular pharmacology and therapeutics*, 24(6), 509–520. <https://doi.org/10.1177/1074248419861437>.
- Lorenzon Dos Santos, J., Quadros, A. S., Weschenfelder, C., Garofallo, S. B., & Marcadenti, A. (2020). Oxidative Stress Biomarkers, Nut-Related Antioxidants, and Cardiovascular Disease. *Nutrients*, 12(3), 682. <https://doi.org/10.3390/nu12030682>.
- Bobescu, E., Marceanu, L. G., Dima, L., Balan, A., Strempel, C. G., & Covaci, A. (2021). Trimetazidine Therapy in Coronary Artery Disease: The Impact on Oxidative Stress, Inflammation, Endothelial Dysfunction, and Long-Term Prognosis. *American journal of therapeutics*, 28(5), e540–e547. <https://doi.org/10.1097/MJT.0000000000001430>.
- Slimen, I. B., Najar, T., Ghram, A., Dabbebi, H., Ben Mrad, M., & Abdrrabbah, M. (2014). Reactive oxygen species, heat stress and oxidative-induced mitochondrial damage. A review. *International journal of hyperthermia : the official journal of European Society for Hyperthermic Oncology, North American Hyperthermia Group*, 30(7), 513–523. <https://doi.org/10.3109/02656736.2014.971446>.
- Juan, C. A., Pérez de la Lastra, J. M., Plou, F. J., & Pérez-Lebeña, E. (2021). The Chemistry of Reactive Oxygen Species (ROS) Revisited: Outlining Their Role in Biological Macromolecules (DNA, Lipids and Proteins) and Induced Pathologies. *International journal of molecular sciences*, 22(9), 4642. <https://doi.org/10.3390/ijms22094642>.

8. Marín, R., Abad, C., Rojas, D., Chiarello, D. I., & Alejandro, T. G. (2023). Biomarkers of oxidative stress and reproductive complications. *Advances in clinical chemistry*, 113, 157–233. <https://doi.org/10.1016/bs.acc.2022.11.004>.
9. Grochowski, C., Litak, J., Kamieniak, P., & Maciejewski, R. (2018). Oxidative stress in cerebral small vessel disease. Role of reactive species. *Free radical research*, 52(1), 1–13. <https://doi.org/10.1080/10715762.2017.1402304>.
10. Zhang, Z., Huang, Q., Zhao, D., Lian, F., Li, X., & Qi, W. (2023). The impact of oxidative stress-induced mitochondrial dysfunction on diabetic microvascular complications. *Frontiers in endocrinology*, 14, 1112363. <https://doi.org/10.3389/fendo.2023.1112363>.
11. Shahcheraghi, S. H., Salemi, F., Small, S., Syed, S., Salari, F., Alam, W., Cheang, W. S., Saso, L., & Khan, H. (2023). Resveratrol regulates inflammation and improves oxidative stress via Nrf2 signaling pathway: Therapeutic and biotechnological prospects. *Phytotherapy research: PTR*, 37(4), 1590–1605. <https://doi.org/10.1002/ptr.7754>.
12. Reghelin, C. K., Bastos, M. S., de Souza Basso, B., Costa, B. P., Lima, K. G., de Sousa, A. C., Haute, G. V., Diz, F. M., Dias, H. B., Luft, C., Rodrigues, K. F., Garcia, M. C. R., Matzenbacher, L. S., Adami, B. S., Xavier, L. L., Donadio, M. V. F., de Oliveira, J. R., & da Silva Melo, D. A. (2023). Bezafibrate reduces the damage, activation and mechanical properties of lung fibroblast cells induced by hydrogen peroxide. *Naunyn-Schmiedeberg's archives of pharmacology*, 396(12), 3857–3866. <https://doi.org/10.1007/s00210-023-02595-2>.
13. Zhang, G. Z., Liu, M. Q., Chen, H. W., Wu, Z. L., Gao, Y. C., Ma, Z. J., He, X. G., & Kang, X. W. (2021). NF-κB signalling pathways in nucleus pulposus cell function and intervertebral disc degeneration. *Cell proliferation*, 54(7), e13057. <https://doi.org/10.1111/cpr.13057>.
14. Nwachukwu, I. D., & Aluko, R. E. (2019). Structural and functional properties of food protein-derived antioxidant peptides. *Journal of food biochemistry*, 43(1), e12761. <https://doi.org/10.1111/jfbc.12761>.
15. Y., Liu, J., Zhou, Q., Chen, X., Ding, X., & Zhang, X. (2022). Expression of LINC00638 in rheumatoid arthritis patients with damp-heat obstruction syndrome and the regulatory mechanisms for inflammation and oxidative stress. *Journal of Central South University. Medical sciences*, 47(2), 183–193. <https://doi.org/10.11817/j.issn.1672-7347.2022.210376>.
16. Houben JM, Moonen HJ, van Schooten FJ, Hageman GJ. Telomere length assessment: biomarker of chronic oxidative stress? *Free Radic Biol Med*. 2008 Feb 1;44(3):235-46.
17. Tian, R., Zhang, L. N., Zhang, T. T., Pang, H. Y., Chen, L. F., Shen, Z. J., Liu, Z., Fang, Q., & Zhang, S. Y. (2017). Association Between Oxidative Stress and Peripheral Leukocyte Telomere Length in Patients with Premature Coronary Artery Disease. *Medical science monitor: international medical journal of experimental and clinical research*, 23, 4382–4390. <https://doi.org/10.12659/msm.902106>.
18. Werner, C., Gensch, C., Pöss, J., Haendeler, J., Böhm, M., & Laufs, U. (2011). Pioglitazone activates aortic telomerase and prevents stress-induced endothelial apoptosis. *Atherosclerosis*, 216(1), 23–34. <https://doi.org/10.1016/j.atherosclerosis.2011.02.011>.
19. Zhang, W., Hui, R., & Yang, S. (2014). Telomeres, cardiovascular aging, and potential intervention for cellular senescence. *Science China. Life sciences*, 57(8), 858–862. <https://doi.org/10.1007/s11427-014-4700-8>.
20. Minamino, T., & Komuro, I. (2002). Role of telomere in endothelial dysfunction in atherosclerosis. *Current opinion in lipidology*, 13(5), 537–543. <https://doi.org/10.1097/00041433-200210000-00010>.
21. Smith, E. M., Pendlebury, D. F., & Nandakumar, J. (2020). Structural biology of telomeres and telomerase. *Cellular and molecular life sciences : CMLS*, 77(1), 61–79. <https://doi.org/10.1007/s00018-019-03369-x>
22. Martínez, P., & Blasco, M. A. (2015). Replicating through telomeres: a means to an

end. *Trends in biochemical sciences*, 40(9), 504–515. <https://doi.org/10.1016/j.tibs.2015.06.003>.

23. Giardini, M. A., Segatto, M., da Silva, M. S., Nunes, V. S., & Cano, M. I. (2014). Telomere and telomerase biology. *Progress in molecular biology and translational science*, 125, 1–40. <https://doi.org/10.1016/B978-0-12-397898-1.00001-3>.

24. Revy, P., Kannengiesser, C., & Bertuch, A. A. (2023). Genetics of human telomere biology disorders. *Nature reviews. Genetics*, 24(2), 86–108. <https://doi.org/10.1038/s41576-022-00527-z>.

25. Chen, B., Yan, Y., Wang, H., & Xu, J. (2023). Association between genetically determined telomere length and health-related outcomes: A systematic review and meta-analysis of Mendelian randomization studies. *Aging cell*, 22(7), e13874. <https://doi.org/10.1111/acel.13874>.

26. Nassour, J., Aguiar, L. G., Correia, A., Schmidt, T. T., Mainz, L., Przetocka, S., Haggblom, C., Tadepalle, N., Williams, A., Shokhirev, M. N., Akincilar, S. C., Tergaonkar, V., Shadel, G. S., & Karlseder, J. (2023). Telomere-to-mitochondria signalling by ZBP1 mediates replicative crisis. *Nature*, 614(7949), 767–773. <https://doi.org/10.1038/s41586-023-05710-8>.

27. Revy, P., Kannengiesser, C., & Bertuch, A. A. (2023). Genetics of human telomere biology disorders. *Nature reviews. Genetics*, 24(2), 86–108. <https://doi.org/10.1038/s41576-022-00527-z>.

28. Robinson, L. G., Jr, Kalmbach, K., Sumerfield, O., Noman, W., Wang, F., Liu, L., & Keefe, D. L. (2024). Telomere dynamics and reproduction. *Fertility and sterility*, 121(1), 4–11. <https://doi.org/10.1016/j.fertnstert.2023.11.012>.

29. Allaire, P., He, J., Mayer, J., Moat, L., Gerstenberger, P., Wilhorn, R., Strutz, S., Kim, D. S. L., Zeng, C., Cox, N., Shay, J. W., Denny, J., Bastarache, L., & Hebring, S. (2023). Genetic and clinical determinants of telomere length. *HGG advances*, 4(3), 100201. <https://doi.org/10.1016/j.xhgg.2023.100201>.

30. Bloom, S. I., Liu, Y., Tucker, J. R., Islam, M. T., Machin, D. R., Abdeahad, H., Thomas, T. G., Bramwell, R. C., Lesniewski, L. A., & Donato, A. J. (2023). Endothelial cell telomere dysfunction induces senescence and results in vascular and metabolic impairments. *Aging cell*, 22(8), e13875. <https://doi.org/10.1111/acel.13875>.

31. Aubert, G., & Lansdorp, P. M. (2008). Telomeres and aging. *Physiological reviews*, 88(2), 557–579. <https://doi.org/10.1152/physrev.00026.2007>.

32. Gao, X., Yu, X., Zhang, C., Wang, Y., Sun, Y., Sun, H., Zhang, H., Shi, Y., & He, X. (2022). Telomeres and Mitochondrial Metabolism: Implications for Cellular Senescence and Age-related Diseases. *Stem cell reviews and reports*, 18(7), 2315–2327. <https://doi.org/10.1007/s12015-022-10370-8>.

33. Shoeb, M., Meier, H. C. S., & Antonini, J. M. (2021). Telomeres in toxicology: Occupational health. *Pharmacology & therapeutics*, 220, 107742. <https://doi.org/10.1016/j.pharmthera.2020.107742>.

34. Khan, S., Chuturgoon, A. A., & Naidoo, D. P. (2012). Telomeres and atherosclerosis. *Cardiovascular journal of Africa*, 23(10), 563–571. <https://doi.org/10.5830/CVJA-2012-056>.

35. Lin, J., & Epel, E. (2022). Stress and telomere shortening: Insights from cellular mechanisms. *Ageing research reviews*, 73, 101507. <https://doi.org/10.1016/j.arr.2021.101507>.

36. Schuermans, A., Nakao, T., Uddin, M. M., Hornsby, W., Ganesh, S., Shadyab, A. H., Liu, S., Haring, B., Shufelt, C. L., Taub, M. A., Mathias, R. A., Kooperberg, C., Reiner, A. P., Bick, A. G., Manson, J. E., Natarajan, P., & Honigberg, M. C. (2023). Age at Menopause, Leukocyte Telomere Length, and Coronary Artery Disease in Postmenopausal Women. *Circulation research*, 133(5), 376–386. <https://doi.org/10.1161/CIRCRESAHA.123.322984>.

37. Chen, B., Yan, Y., Wang, H., & Xu, J. (2023). Association between genetically determined telomere length and health-related outcomes: A systematic review and meta-analysis of Mendelian randomization studies. *Aging cell*, 22(7), e13874. <https://doi.org/10.1111/acel.13874>.

A systematic review and meta-analysis of Mendelian randomization studies. *Aging cell*, 22(7), e13874. <https://doi.org/10.1111/acel.13874>.

38. Zimnitskaya, O. V., Petrova, M. M., Lareva, N. V., Cherniaeva, M. S., Al-Zamil, M., Ivanova, A. E., & Shnayder, N. A. (2022). Leukocyte Telomere Length as a Molecular Biomarker of Coronary Heart Disease. *Genes*, 13(7), 1234. <https://doi.org/10.3390/genes13071234>.

39. Yin, H., & Pickering, J. G. (2023). Telomere Length: Implications for Atherogenesis. *Current atherosclerosis reports*, 25(3), 95–103. <https://doi.org/10.1007/s11883-023-01082-6>.

40. Ojeda-Rodriguez, A., Rangel-Zuñiga, O. A., Arenas-de Larriva, A. P., Gutierrez-Mariscal, F. M., Torres-Peña, J. D., Romero-Cabrera, J. L., Podadera-Herreros, A., García-Fernandez, H., Porras-Pérez, E., Luque, R. M., Kales, S. N., Perez-Martinez, P., Delgado-Lista, J., Yubero-Serrano, E. M., & Lopez-Miranda, J. (2024). Telomere length as biomarker of nutritional therapy for prevention of type 2 diabetes mellitus development in patients with coronary heart disease: CORDIOPREV randomised controlled trial. *Cardiovascular diabetology*, 23(1), 98. <https://doi.org/10.1186/s12933-024-02175-5>.

41. Codd V., Nelson C.P., Albrecht E., Mangino M., Deelen J., Buxton J.L., Hottenga J.J., Fischer K., Esko T., Surakka I., et al. Identification of seven loci affecting mean telomere length and their association with disease. *Nat. Genet.* 2013;45:422–427.

42. Sethi I, Bhat GR, Singh V, Kumar R, Bhanwer AJ, Bamezai RN, Sharma S, Rai E. Role of telomeres and associated maintenance genes in Type 2 Diabetes Mellitus: A review. *Diabetes Res Clin Pract.* 2016 Dec; 122:92-100.

43. Kuznetsova T, Codd VS, Lutgarde T, Gonzalez A, Jin Y, Richart T, van der Harst P, Diez J, Staessen JA, Samani NJ: Association between left ventricular mass and telomere length in a population study. *Am J Epidemiol* 2010, 172:440–450.